

**ANA FLÁVIA SORIANO PEREIRA**

**AVALIAÇÃO *IN VITRO* DO EFEITO BACTERIOSTÁTICO DO PLASMA RICO EM PLAQUETAS (PRP) EM *STAPHYLOCOCCUS AUREUS* SENSÍVEL A METICILINA (MSSA) E *STAPHYLOCOCCUS AUREUS* RESISTENTES A METICILINA (MRSA) PELO TESTE KILL CURVE ASSAY**

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PELO TESTE *KILL CURVE ASSAY***

Dissertação apresentada à Pró-Reitoria de Pesquisa e Pós-Graduação, Universidade do Oeste Paulista, como parte dos requisitos para obtenção do título de Mestre em Ciência Animal – Área de concentração: Fisiopatologia Animal.

Orientadora:  
Profa. Dra. Rosa Maria Barilli Nogueira  
Co-Orientador:  
Dr. Rodrigo Costa Silva

636.089      Pereira, Ana Flávia Soriano.  
P246a           Avaliação *in vitro* do efeito bacteriostático do plasma  
rico em plaquetas (PRP) em *staphylococcus aureus*  
sensível a meticilina (MSSA) e *staphylococcus aureus*  
resistentes a meticilina (MRSA) pelo teste *kill curve assay*  
/ Ana Flávia Soriano Pereira. – Presidente Prudente,  
2021.  
64f.: il.

Dissertação (Mestrado em Ciência Animal) -  
Universidade do Oeste Paulista – Unoeste, Presidente  
Prudente, SP, 2021.

Bibliografia.

Orientador: Rosa Maria Barilli Nogueira  
Co-Orientador: Rodrigo Costa Silva

1. Estafilococos. 2. Resistência Bacteriana. 3.  
Interação plaqueta-leucócito. 4. Conteúdo plaquetário. I.  
Título.

Catalogação na Fonte: Michele Mologni – CRB 8 -6204

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Presidente Prudente, 04 de novembro de 2021.

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## **DEDICATÓRIA**

Dedico à minha família e noivo, que se dedicaram ao máximo para que eu conquistasse todos os meus sonhos até agora e sempre foram pilar em minha jornada me mantendo em pé.

## **AGRADECIMENTOS**

Primeiramente agradeço a Deus, pela força e coragem durante esta caminhada.

À minha mãe Eliete Soriano Pereira e irmão Luiz Guilherme Soriano Pereira assim como meu noivo Lucas Martins Pacheco que, com muito carinho, apoio, compreensão e colaboração acreditaram em mim, entenderam minha ausência e me incentivaram a continuar diante de todas as dificuldades.

E em especial com todo meu amor, carinho e saudade (in memorian) ao meu pai amado Aldo Pereira Junior que tanto me apoiou e sonhou com o término desse trabalho, e meu irmão Luiz Eduardo Soriano Pereira que do céu celebrou minha vitória como sempre fez aqui na terra.

A minha orientadora, Profa. Dra. Rosa Maria Barilli Nogueira, uma grande mulher, que me aceitou como sua orientanda e dividiu comigo seus projetos e sonhos. Não tenho como agradecer tudo o que fez por mim. Você é uma pessoa iluminada, daquelas diferentes que apareçam e marcam nossas vidas. Obrigada por estar ao meu lado nos melhores e piores momentos, sempre me incentivando e compartilhando comigo dos meus choros e sorrisos. E no fim de cada conversando me motivando com a seguinte frase “Tenho certeza que você consegue e dá conta Ana”. Obrigada por tudo.

Ao meu co-orientador, Prof. Dr. Rodrigo Costa Silva, que tanto colaborou para a realização deste trabalho ajudando na parte experimental, nas análises do projeto, assim como em todas as etapas do trabalho não medindo esforços para nos ajudar, você foi fundamental para a concretização do trabalho obrigada pela ajuda e disponibilização de tempo para toda realização do projeto.

A Unoeste, pela oportunidade de estudo no Mestrado e por todo o apoio dos profissionais que nos atendem com dedicação, especialmente Keid Ribeiro Kruger.

A Banca examinadora por ter aceito o convite de participar deste momento tão importante.

À CAPES pelo incentivo financeiro concedido: “O presente trabalho foi realizado com apoio da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – (Brasil) CAPES.

Obrigada a todos!

*“A tarefa não é tanto ver aquilo que ninguém viu, mas pensar o que ninguém ainda pensou sobre aquilo que todo mundo vê.”*

*(Arthur Schopenhauer)*

## RESUMO

### Avaliação *in vitro* do efeito bacteriostático do plasma rico em plaquetas (PRP) em *Staphylococcus aureus* sensível a meticilina (MSSA) e *Staphylococcus aureus* resistentes a meticilina (MRSA) pelo teste *kill curve assay*

Infecções estafilocócicas apresentam relevância pela frequência que ocorrem e gravidade. *Staphylococcus aureus* (*S. aureus*) é uma bactéria Gram-positiva relevante por comumente causar infecções, pelo aumento dos casos de resistência, se desenvolvendo rapidamente e representando um grave problema de saúde pública no mundo. As plaquetas têm destaque na participação em processos infecciosos e inflamatórios pela interação com o patógeno, em especial as bactérias e as células de defesa podem ser obtidas em maior concentração no plasma rico em plaquetas (PRP), um concentrado de sangue com diferentes concentrações de fatores de crescimento, plaquetas e leucócitos, e que vem sendo utilizado de forma multidisciplinar, em regeneração tecidual e com propriedades antimicrobianas. Diante da hipótese que as plaquetas são as responsáveis pelo efeito bacteriostático do PRP frente ao *S. aureus*, objetivou-se avaliar o efeito bacteriostático *in vitro* de plaquetas caninas sobre *S. aureus* resistente à meticilina (MRSA) e *S. aureus* sensível a meticilina (MSSA) utilizando-se compostos contendo diferentes concentrações de plaquetas e leucócitos caninos. Desenhou-se estudo experimental controlado randomizado em bloco, comparando-se o efeito bacteriostático das plaquetas, em 6 grupos experimentais frente MRSA (G1-G6) e MSSA (G7-G12): G1 e G7, salina; G2 e G8, controle negativo; G3 e G9, meio de cultura; G4 e G10, PRP, G5 e G11, plasma puro rico em plaquetas (P-PRP), e G6 e G12, leucócitos isolados (WBC). Utilizou-se o modelo de teste *kill curve assay* para avaliar a multiplicação bacteriana por contagem de unidades formadoras de colônia (UFC/mL) em meio de cultura Mueller-Hinton (MHB), após incubação das bactérias com seus respectivos tratamentos em caldo MHB por 0h, 1h e 2 horas. PRP apresentou taxa de contaminação por leucócitos de 1:12,59 plaquetas, não observada no P-PRP, permitindo a avaliação individualizada das plaquetas sobre as bactérias. Para ambas as bactérias, observou-se que leucócitos (WBC) e a interação plaqueta:leucócito induziram uma redução na multiplicação bacteriana ao longo do experimento, não havendo diferença significativa entre os time points ( $p>0,05$ ). Os efeitos dos tratamentos a cada momento e comparativo ao grupo controle basal (G1 e G7), observou-se comportamento similar para ambas as bactérias na primeira hora de incubação, com redução da multiplicação bacteriana nos grupos tratados somente com WBC (G5 = -2,86x; G11 = -8,79 x), seguido pelo tratamento com PRP (G4 = -1,69x; G10 = -3,96x), fato este observado na segunda hora de incubação, porém com redução da multiplicação bacteriana no G5 (-1,07x) e G11 (-2,36x), e uma intensa redução no grupo PRP somente para MSSA (G10 = -8,03x), fato este não observado para MRSA (G5 = 1,33x). Interessantemente, as plaquetas isoladas estimularam a multiplicação bacteriana tanto na primeira hora de incubação (G6 = 1,53x; G12 = 1,24x), quanto na segunda hora (G12 = 1,50x), com exceção do G6 (-1,16x). Este efeito pode ser devido a interação física plaqueta:leucócito, como do conteúdo dos grânulos plaquetários e leucócitos. Assim, as plaquetas desempenham papel importante na inibição/redução da multiplicação bacteriana *in vitro* de MRSA e MSSA quando na presença de leucócitos, enfatizando a interação sinérgica principalmente nos primeiros momentos de exposição ao patógeno.

**Palavras chaves:** Estafilococos. Resistência bacteriana. Interação plaqueta-leucócito. Conteúdo plaquetário.

## ABSTRACT

### ***In vitro evaluation of the bacteriostatic effect of platelet-rich plasma (PRP) on methicillin-sensitive *Staphylococcus aureus* (MSSA) and methicillin-resistant *Staphylococcus aureus* (MRSA) by the kill curve assay***

Staphylococcal infections are relevant for their frequency and severity. *Staphylococcus aureus* (*S. aureus*) is a relevant Gram-positive bacterium because it commonly causes infections, because of the increase in resistance cases, it develops quickly and represents a serious public health problem in the world. Platelets stand out in their participation in infectious and inflammatory processes due to their interaction with the pathogen, in particular bacteria and defense cells can be obtained in higher concentration in platelet-rich plasma (PRP), a concentrate of blood with different concentrations of factors of growth, platelets and leukocytes, and that has been used in a multidisciplinary way, in tissue regeneration and with antimicrobial properties. Given the hypothesis that platelets are responsible for the bacteriostatic effect of PRP against *S. aureus*, this study aimed to evaluate the in vitro bacteriostatic effect of canine platelets on methicillin-resistant *S. aureus* (MRSA) and methicillin-sensitive *S. aureus* (MSSA) using compounds containing different concentrations of platelets and canine leukocytes. A randomized block experimental study was designed, comparing the bacteriostatic effect of platelets, in 6 experimental groups against MRSA (G1-G6) and MSSA (G7-G12): G1 and G7, saline; G2 and G8, negative control; G3 and G9, culture medium; G4 and G10, PRP, G5 and G11, pure platelet-rich plasma (P-PRP), and G6 and G12, isolated leukocytes (WBC). The kill curve assay model was used to assess bacterial multiplication by counting colony-forming units (CFU/mL) in Mueller-Hinton (MHB) culture medium, after incubation of the bacteria with their respective treatments in MHB broth by 0h, 1h and 2 hours. PRP had a leukocyte contamination rate of 1:12.59 platelets, not observed in P-PRP, allowing the individualized evaluation of platelets on bacteria. For both bacteria, it was observed that leukocytes (WBC) and platelet:leukocyte interaction induced a reduction in bacterial multiplication throughout the experiment, with no significant difference between time points ( $p>0.05$ ). The effect of treatments at each time point and compared to the basal control group (G1 and G7), a similar behavior was observed for both bacteria in the first hour of incubation, with a reduction in bacterial multiplication in the groups treated only with WBC (G5 = -2 .86x; G11 = -8.79x), followed by treatment with PRP (G4 = -1.69x; G10 = -3.96x), a fact observed in the second hour of incubation, but with reduced bacterial multiplication in G5 (-1.07x) and G11 (-2.36x), and an intense reduction in the PRP group only for MSSA (G10 = -8.03x), a fact not observed for MRSA (G5 = 1.33x). Interestingly, the isolated platelets stimulated bacterial multiplication both in the first hour of incubation (G6 = 1.53x; G12 = 1.24x) and in the second hour (G12 = 1.50x), with the exception of G6 (-1.16x). This effect may be due to physical platelet:leukocyte interaction, such as the content of platelet granules and leukocytes. Thus, platelets play an important role in the inhibition/reduction of bacterial multiplication in vitro of MRSA and MSSA when in the presence of leukocytes, emphasizing the synergistic interaction, especially in the first moments of exposure to the pathogen.

**Keywords:** Staphylococci. Bacterial Resistance. Platelet-leukocyte interaction. Platelet content.

## **LISTA DE SIGLAS**

*S. aureus* – *Staphylococcus aureus*

MRSA – *Staphylococcus aureus* resistente a meticilina

β-lactânicos – Beta-lactânicos

MSSA – *Staphylococcus aureus* sensível a meticilina

α-grânulos – Grânulos alfa

PMPs – Proteínas/peptídeos microbicidas plaquetários

PAMs – Peptídeos antimicrobianos

PRP – Plasma rico em plaquetas

ANOVA – Análise de variância

UFC – Unidades formadoras de colônia

CEUA – Comitê de Ética no Uso de Animais em Experimentação

Kg – Quilograma

SRD – Sem raça definida

mL – Mililitro

% – Porcentagem

µg – Micrograma

≥ – Maior ou igual

mm – Milímetro

°C – Graus Celsius

PLT – Contagem total de plaquetas

MPV fL – Volume plaquetário médio

PDW% - Amplitude de distribuição plaquetária

WBC – Contagem total de leucócitos

RBC – Contagem total de eritrócitos

HCT – Volume globular

Hb – Hemoglobina

x – Vezes

mol/L – Mol por litro

g – gramas

µL – Microlitro

GC – Gluconato de cálcio

PRG – Gel rico em plaquetas

h – Horas

UFC/mL – Unidades formadoras de colônia por mililitro

SST – Solução salina tamponada de fosfatos

rpm – Rotação por minuto

## SUMÁRIO

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**ARTIGO ORIGINAL**

**Avaliação *in vitro* do efeito bacteriostático do plasma rico em plaquetas (PRP) em *Staphylococcus aureus* sensível a meticilina (MSSA) e *Staphylococcus aureus* resistentes a meticilina (MRSA) pelo teste *kill curve assay***

***In vitro evaluation of the bacteriostatic effect of platelet-rich plasma (PRP) on methicillin-sensitive *Staphylococcus aureus* (MSSA) and methicillin-resistant *Staphylococcus aureus* (MRSA) by the kill curve assay***

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## RESUMO

Infecções estafilocócicas apresentam relevância pela frequência e gravidade. *Staphylococcus aureu* (*S. aureus*), bactéria gram-positiva, relevante em infecções, tem seu tratamento comprometido devido casos de resistência. Plaquetas, em processos infecciosos e inflamatórios, interagem com o patógeno e são obtidas em maior concentração no plasma rico em plaquetas (PRP), destacando-se pelo uso com propriedades antimicrobianas. Foi avaliado o efeito bactericida *in vitro* de plaquetas caninas sobre *S. aureus* resistente à meticilina (MRSA) e *S. aureus* sensível a meticilina (MSSA) utilizando-se diferentes compostos e concentrações de plaquetas e leucócitos caninos. Comparou-se o efeito bactericida das plaquetas, em 6 grupos experimentais frente MRSA (G1-G6) e MSSA (G7-G12): G1xG7 salina; G2xG8 controle negativo; G3xG9 meio de cultura; G4xG10 PRP; G5xG11 plasma puro rico em plaquetas (P-PRP); G6xG12 leucócitos isolados (WBC). Utilizou-se kill curve assay para avaliar multiplicação bacteriana por contagem de unidades formadoras de colônia (UFC/mL). PRP apresentou taxa de contaminação por leucócitos, não observada no P-PRP, permitindo avaliação individualizada das plaquetas sobre as bactérias. Efeito bactericida foi avaliado por kill curve assay, com base na contagem de UFC/mL, em diferentes *time points*. Para ambos, observou-se que leucócitos e interação plaqueta:leucócito induziram redução na multiplicação bacteriana. Analisando o efeito dos tratamentos, observou-se comportamento similar para ambos na 1<sup>ª</sup> hora de incubação, com redução da multiplicação bacteriana mais intensa nos grupos tratados somente com WBC, seguido pelo tratamento com PRP, fato observado também na 2<sup>ª</sup> hora. Plaquetas isoladas estimularam multiplicação bacteriana tanto na primeira hora de incubação, quanto na segunda, o que pode ser devido interação física plaqueta:leucócito e conteúdo dos grânulos plaquetários e leucócitos. Assim, plaquetas desempenham papel importante na inibição/redução da multiplicação bacteriana *in vitro* de MRSA e MSSA quando na presença de leucócitos, enfatizando interação sinérgica principalmente nos primeiros momentos de exposição ao patógeno.

**Palavras chaves:** Estafilococos. Resistência bacteriana. Interação plaqueta-leucócito. Conteúdo Plaquetário.

## 1. INTRODUÇÃO

A resistência antimicrobiana é a capacidade dos microrganismos de resistir ao efeito dos medicamentos (antibióticos), uma ameaça séria que vem aumentando constantemente e se tornando um crescente problema de saúde no mundo todo, gerando profundas implicações para a saúde pública. Na fase em que nos encontramos atualmente, até infecções simples se tornam intratáveis e, com isso, o surgimento de cepas resistentes tem seu crescimento acelerado [1]. A pesquisa neste campo está focada em encontrar novos agentes e estratégias para combater infecções e reduzir o tempo de tratamento [2].

*Staphylococcus aureus* (*S. aureus*) é um dos mais importantes e letais patógenos bacterianos identificados desde o início do século XX. Bactéria gram-positiva, clinicamente relevante, sendo o isolado bacteriano mais comumente observado em várias infecções e associado a maiores taxas de falha do tratamento, devido a capacidade de formar biofilmes [3-5]. Por definição prática, *Staphylococcus aureus* resistente a meticilina (MRSA) apresenta resistência a todos os antibióticos betalactâmicos ( $\beta$ -lactâmicos) enquanto *Staphylococcus aureus* sensível a meticilina (MSSA) apresenta sensibilidade a esta classe de antibióticos [6]. A primeira linha de escolha para o tratamento de infecções complicadas por MRSA é o antibiótico vancomicina, porém devido aos seus efeitos colaterais buscam-se novas alternativas para terapias eficazes e seguras [7].

As plaquetas, como já observado na literatura, desempenham papel fundamental na defesa do hospedeiro contra a infecções [8]. Elas têm a capacidade de liberar proteínas e fatores de crescimento e, segundo descobertas mais recentes, também podem liberar agentes imunomoduladores com atividade antimicrobiana [9-10]. Estes fragmentos dos megacariócitos (células precursoras das plaquetas e presentes na medula óssea) interagem com o patógeno e células inflamatórias, acumulando-se no local do dano endotelial causado pela colonização microbiana e desempenhando papel defensivo natural na luta contra as infecções. As plaquetas possuem diversas organelas e vacúolos no seu interior, dentre eles os grânulos alfa ( $\alpha$ -grânulos) e grânulos denso. Dentre estes, os grânulos alfa são ricos em fatores de crescimento e citocinas, se destacam por estimular a resposta inflamatória, recrutando células do sistema imunológico para o local da lesão para atacar os patógenos. Quando expostas às bactérias, as plaquetas participam da co-adesão bacteriana, resultando no sequestro bacteriano e na fagocitose das bactérias. Também auxiliam os neutrófilos na criação de interações célula-célula

com células endoteliais e leucócitos. Desta forma, participam da resposta imune inata do organismo. Os grânulos alfa também contêm moléculas conhecidas como proteínas/peptídeos microbicidas plaquetários (PMPs), que, quando liberados, têm um efeito antimicrobiano [11-14].

As PMPs são consideradas a primeira linha de defesa contra microrganismos invasores, pois suas moléculas liberadas após a ativação plaquetária são capazes de recrutar células e modular múltiplos processos fisiológicos [15-16]. Outra classe muito promissora para o tratamento são os peptídeos antimicrobianos (PAMs), compostos de aminoácidos produzidos pelas classes de organismos multicelulares como parte da resposta imune nata. Também exercem atividade inibidora intracelular. A vantagem é que são menos propensos a induzir resistência por possuírem múltiplos alvos e raramente interagirem com um receptor específico [1-17]. Olhar para os vários concentrados de plaquetas apenas como um agente antimicrobiano limitaria seu uso. Eles devem ser vistos como agentes com efeitos regenerativos e antimicrobianos e assim, podem ser designados como um dos melhores tratamentos para infecções complicadas [9].

As plaquetas podem ser trabalhadas externamente ao organismo de modo a explorar suas atividades e substâncias. Para tal, atualmente compostos ricos em plaquetas (plasma rico em plaquetas-PRP) têm sido obtido mediante diversos protocolos para diferentes espécies. O PRP é uma porção do plasma autólogo e possui uma concentração de plaquetas acima da linha basal, sendo assim rico em fatores de crescimento e citocinas que estão envolvidos em uma resposta inflamatória [3-11-18]. Com isso, o tratamento com o PRP, pode gerar então efeito antimicrobiano, também [11]. Atualmente o PRP vem sendo cada vez mais utilizado em múltiplas aplicações clínicas, mostrando resultados promissores principalmente em regeneração tecidual [19-20]. Pode ser utilizado também como biomaterial, e foi aplicado pela primeira vez como uma "cola" em cirurgias na década de 1970 (essencialmente idêntico ao da cola de fibrina atual) [21]. O PRP pode funcionar então como um complemento à terapia antibiótica convencional ou como uma alternativa devido ao risco potencialmente menor de resistência aos medicamentos [15-22].

Além das plaquetas, outras células encontradas no PRP também podem contribuir para um efeito antimicrobiano, como os leucócitos, a composição pode ser alterada para incluir mais ou menos determinada células específicas durante o processamento [2-11]. Alguns estudos com PRP rico em leucócitos mostram que a

resposta antibacteriana pode ser mais significativa, possivelmente devido à agregação leucócito-plaquetas causando um aumento da resposta inflamatória [19-23].

Apesar do grande potencial de aplicabilidade, o uso terapêutico do PRP como uma alternativa clínica tornou-se difícil, devido à falta de estudos relacionados com a padronização das técnicas e/ou insuficiente descrição dos procedimentos adotados, e a possibilidade de incluir diferentes concentrações de substâncias dificulta as conclusões de como se dá suas propriedades antibacterianas [15-24]. Por essas razões e observações, hipotetizamos que, além de suas propriedades promotoras de cura, pode possuir propriedades antimicrobianas. Ao utilizá-lo como agente bacteriostático temos a vantagem de o mesmo apresentar menor potencial de desenvolver resistência bacteriana em comparação com antibioticoterapia convencional.

Frente a hipótese de que o uso do PRP pode ser um tratamento alternativo a infecções complicadas por MSSA e MRSA, as plaquetas são as responsáveis pelo efeito bacteriostático do PRP frente a bactéria *S. aureus*, o presente estudo objetivou avaliar o efeito bacteriostático *in vitro* de plaquetas caninas em *Staphylococcus aureus* resistente a meticilina (MRSA) e *Staphylococcus aureus* sensível a meticilina (MSSA) por meio do teste kill curve assay utilizando diferentes compostos contendo concentrações diferentes de plaquetas e leucócitos caninos.

## 2. MÉTODOS

Todos os experimentos foram realizados nos laboratórios do Hospital Veterinário, da Faculdade de Ciências Agrárias, Universidade do Oeste Paulista-Unoeste. As coletas de sangue foram realizadas nas dependências do canil do Hospital Veterinário, foram feitas duas coletas com intervalo de tempo de 30 dias. O processamento das amostras de sangue, hemograma, contagem de células e plaquetas, e processamento das suspensões utilizadas como tratamento no presente projeto foi realizado no Laboratório de Análises Clínicas. O processamento das amostras microbiológicas e exposição dos tratamentos às bactérias utilizadas no estudo foram realizadas no Laboratório de Microbiologia Veterinária.

### 2.1 Delineamento amostral

Todos os experimentos foram realizados utilizando um desenho experimental controlado randomizado em bloco. O tamanho amostral proposto no presente estudo foi calculado utilizando-se o método de equação de recursos (resource

equation) com base na lei dos rendimentos decrescentes (law of diminishing returns), descrito por Festing [25]. Resumidamente, o tamanho amostral apropriado foi determinado pelo número de graus de liberdade para o erro na análise de variância (ANOVA) ou teste t, utilizando a fórmula:  $E = N - T - B$ , onde E, N, T, e B correspondem ao erro, total de observações, tratamentos e blocos (replicatas) utilizados no ANOVA, respectivamente. Para este cálculo, o possível erro, E, deve estar entre 10 e 20. Assim, considerando cada tubo utilizando o teste kill curve assay como uma unidade experimental e cada tratamento (PRP, WBC, P-PRP, e os três controles) sendo realizado em duplicata, onde cada replicata da duplicata foi plaqueada somente uma vez nas diluições 10-1 e 10-2, foi necessária a repetição do experimento duas vezes (portanto, duas replicatas ou dois blocos), com cada diferente momento de avaliação (time points) considerado como um momento de unidade experimental. Foi realizada a leitura somente da diluição que apresentou menor quantidade de unidades formadoras de colônias (UFC), porém acima de 30 UFCs. Assim, o cálculo com duas replicatas é:  $E = 18 - 5 - 1 = 12$  (para cada bactéria), porque foram 18 observações totais (uma em cada *timepoint*), seis tratamentos, e dois blocos, para cada bactéria testada.

O presente trabalho foi aprovado pelo Comitê de Ética no Uso de Animais em Experimentação (CEUA), da Universidade do Oeste Paulista (Unoeste), sob o protocolo no 5969 (em anexo).

## 2.2 Animais

Dois animais, da espécie canina (*Canis familiaris*), uma fêmea e um macho, adultos, acima de 25Kg, sem raça definida (SRD), hígidos, mantidos sob cuidados médico veterinário no canil do Hospital Veterinário da Unoeste, foram selecionados e utilizados para as coletas de sangue, após jejum de 12 horas, durante o projeto. Os critérios de inclusão foram, ausência de exposição a qualquer tipo de medicação ou vacinação por até dois meses antes do experimento, alimentação controlada com ração comercial, ausência de doenças hematológicas desde o nascimento, resultado negativo para pesquisa de hematozoários. O estado de saúde destes animais foi confirmado através de exames físicos e rotina de triagem sanguínea, com hemograma completo, incluindo pesquisa de hematozoários, além de bioquímica sérica no Analisador Bioquímico Cobas CIII (Roche, Brasil).

Para o experimento em si, amostras de sangue foram coletadas em 12 tubos vacutainer de 4 mL contendo anticoagulante citrato de sódio 3,2% (obtenção de

PRP, WBC e P-PRP) um tubo vacutainer contendo anticoagulante EDTA (hemograma e contagem basal de plaquetas), e um tubo vacutainer seco (triagem para o estado de saúde), a partir da punção da veia jugular (10-20 mL). As amostras foram coletadas somente no momento do processamento e homogeneizadas por inversão, por meio de vacutainer, para se evitar alteração nos testes hematológicos, além de ativação e agregação plaquetária. Todas as amostras foram processadas no Laboratório de Análises Clínicas, do Hospital Veterinário, Unoeste, previamente ao momento da exposição às bactérias.

### **2.3 Cepas bacterianas**

Foram utilizadas duas cepas da bactéria Gram-positiva, *Staphylococcus aureus* (*S. aureus*), isoladas de amostras clínicas na rotina do Laboratório de Microbiobiologia Veterinária, do Hospital Veterinário, da Faculdade de Ciências Agrárias, Unoeste, após classificação morfológica e bioquímica, e mantidas refrigeradas em estoque no laboratório. As cepas apresentaram resultados positivo para catalase, DNase e coagulase, resultado negativo para D-teste, além de resistência ao antimicrobiano polimixina B 300 $\mu$ g. Adicionalmente, ambas as cepas foram testadas para resistência/sensibilidade a meticilina ( $\beta$ -lactâmicos) mediante a exposição da bactéria ao antimicrobiano cefoxitina 30 $\mu$ g em teste de disco difusão, sendo então classificadas como *Staphylococcus aureus* resistente a meticilina (MRSA) quando presente halo de inibição  $\geq$  22 mm, ou *Staphylococcus aureus* sensível a meticilina (MSSA) quando presente halo de inibição  $<$  22 mm, de acordo com o Manual de teste de sensibilidade a antimicrobianos do Clinical & Laboratory Standards Institute (CLSI/2019) [26].

Ambas as cepas bacterianas foram mantidas em estoque, congeladas, no laboratório. Ao iniciar o projeto, foram semeadas em meio de cultura ágar sangue ovino 5% (BPA-5%) e incubadas a 37°C, 24-48 horas, em estufa de cultura bacteriológica ECB-2 (Olidef, Brasil). Após crescimento, realizou-se novo antibiograma, testando-se diferentes princípios ativos  $\beta$ -lactâmicos para confirmação. Uma vez confirmados os perfis MRSA e MSSA, ocorreu o início das coletas de sangue para o desenvolvimento do estudo. Todos os procedimentos microbiológicos foram realizados no interior de Capela Fluxo Laminar 410 (Pachane, Brasil) e ao redor de bico de Bunsen.

## 2.4 Obtenção de PRP, P-PRP e WBC

Hemograma do animal foi realizado previamente no início do experimento, utilizando-se contador automático Sysmex Poch Diff 100iV-Roche (Sysmex do Brasil, Brasil). Foram determinadas a contagem total de plaquetas (PLT), volume plaquetário médio (MPV fL), amplitude de distribuição plaquetária (PDW%), contagem total de leucócitos (WBC), contagem total de eritrócitos (RBC), volume globular (HCT), e hemoglobina (Hb, fL). A porcentagem de rendimento plaquetário foi calculada pela fórmula: [(número de plaquetas na suspensão analisada / número de plaquetas no sangue total) x 100]. Previamente ao experimento, um piloto foi realizado para validar a contagem plaquetária no contador automático Sysmex Poch Diff 100iV-Roche com amostras contadas manualmente com hemocitômetro (câmara de Neubauer). Adicionalmente, esfregaço sanguíneo foi realizado a partir da amostra de sangue coletada em vacutainer contendo EDTA para determinação dos valores absolutos de cada população leucocitária, confirmação da ausência de hemoparasitas, além da confirmação de ausência de agregados plaquetários. As lâminas foram coradas em corante panótico rápido (Laborclin, Brasil) e visualizadas em microscópio binocular Eclipse E200 (Nikon, Brasil). Este procedimento foi realizado posteriormente em outras etapas do protocolo de obtenção de PRP, P-PRP e WBC para confirmação de ausência de agregação plaquetária.

### 2.4.1 Plasma rico em plaquetas (PRP)

A obtenção de PRP foi realizada após adaptação do protocolo descrito por Vendramin et al. [27]. Amostras de sangue total foram coletadas em tubo contendo anticoagulante citrato de sódio tamponado 3,2% (0,109 mol/L). Os tubos foram centrifugados inicialmente a 200 x g em centrífuga Excelsa Baby 206R, durante 10 minutos, para formação de dois níveis: um superior (sobrenadante) turvo levemente amarelado e um sedimento avermelhado. Toda a fração correspondente ao plasma (sobrenadante), mais 200 µL da fração vermelha, foi transferida para um novo tubo de centrífuga côncico de 15-mL, esterilizado, e novamente centrifugado a 200 x g por 10 minutos. Nesta etapa, formou-se dois níveis distintos: um superior (sobrenadante) turvo levemente amarelado e um sedimento avermelhado. Todo sobrenadante amarelado foi separado em novo tubo de centrífuga crônico de 15-mL, esterilizado, para posterior utilização no experimento. Esta fração foi considerada o PRP. Uma pequena alíquota foi retirada para realização de nova contagem de plaquetas, eritrócitos e leucócitos

automatizada, assim como determinação de agregados plaquetários em esfregaço sanguíneo corado pelo corante panótico rápido. O PRP foi separado para utilização nos experimentos.

#### **2.4.2 Plasma rico em plaquetas puro (P-PRP)**

A obtenção de P-PRP foi realizada após adaptação do protocolo descrito por Vendramin et al. [27] e Mariani et al. [28]. Amostras de sangue total foram coletadas em tubo contendo anticoagulante citrato de sódio tamponado 3,2% (0,109 mol/L). Os tubos foram centrifugados inicialmente a 200 x g em centrífuga Excelsa Baby 206R, durante 10 minutos, para formação de dois níveis: um superior (sobrenadante) turvo levemente amarelado e um sedimento avermelhado. Toda a fração correspondente ao plasma (sobrenadante), mais 200 µL da fração vermelha, foi transferida para um novo tubo de centrífuga cônico de 15-mL, esterilizado, e centrifugado a 400 x g por 10 minutos. Nesta etapa, formou-se uma suspensão sobrenanante turva com coloração levemente esbranquiçada e um sedimento em forma de “botão de fundo” avermelhado. Todo sobrenadante esbranquiçado foi separado em novo tubo de centrífuga crônico de 15-mL, esterilizado, e centrifugado a 643 x g por 10 minutos. Nesta etapa, formou-se um sobrenadante límpido e um sedimento levemente esbranquiçado turvo, contendo as plaquetas. Todo o sobrenadante foi retirado, e o sedimento ressuspendido no próprio sobrenadante removido de modo a se obter a concentração necessária para utilização em todos os respectivos grupos no desafio as bactérias. Esta fração foi considerada o P-PRP. Uma pequena alíquota foi retirada para realização de nova contagem de plaquetas, eritrócitos e leucócitos automatizada, com o objetivo de se verificar a concentração da suspensão em relação a concentração inicial. O P-PRP foi separado para utilização nos experimentos.

#### **2.4.3 Leucócitos isolados (White blood cells, WBC)**

A obtenção de leucócitos isolados (WBC) foi realizada juntamente com o protocolo de obtenção de PRP e, assim, adaptado do protocolo descrito por Vendramin et al. [27]. Desta forma, as amostras de sangue total foram coletadas em tubo contendo anticoagulante citrato de sódio tamponado 3,2% (0,109 mol/L), como para PRP. Os tubos foram centrifugados inicialmente a 200 x g em centrífuga Excelsa Baby 206R, durante 10 minutos, para formação de dois níveis: um superior (sobrenadante) turvo

levemente amarelado e um sedimento avermelhado. Toda a fração correspondente ao plasma (sobrenadante), mais 200 µL da fração vermelha, foi transferida para um novo tubo de centrífuga cônico de 15-mL, esterilizado, e novamente centrifugado a 200 x g por 10 minutos. Nesta etapa, formou-se dois níveis distintos: um superior (sobrenadante) turvo levemente amarelado e um sedimento avermelhado. Todo sobrenadante amarelado foi separado em novo tubo de centrífuga crônico de 15-mL, esterilizado, para posterior utilização no experimento. Para se obter somente WBC, esta suspensão foi induzida a ativação e agregação plaquetária pela agitação forçada em agitador de tubos. A suspensão foi centrifugada a 2000 x g, desprezado o sobrenadante e o sedimento ressuspêndido em solução fisiológica 0,9% estéril. Este procedimento foi repetido mais duas vezes. Esta fração foi determinada como WBC. Uma pequena alíquota foi retirada para realização de nova contagem de plaquetas, eritrócitos e leucócitos automatizada, assim como identificação de plaquetas e leucócitos em esfregaço sanguíneo corado pelo corante panótico rápido. Deste modo, foi obtido a suspensão WBC com alto grau de pureza. O WBC foi separado para utilização nos experimentos.

## 2.5 Ativação plaquetária

A ativação plaquetária foi realizada após adaptação do protocolo descrito por Vendramin et al. [27] e Li [3] nos microtubos utilizados no momento do experimento. Adicionou-se 1 parte de gluconato de cálcio (GC) 10% (9,3 mg/mL) para 4 partes de cada uma das suspensões PRP, WBC, e P-PRP (proporção de 1:4) para induzir a ativação plaquetária e produzir gel (coágulo) rico em plaquetas (PRG) nas três suspensões (Quadro 1). Os volumes foram descritos em proporções visto que as concentrações de plaquetas (PRP e P-PRP) leucócitos (WBC) foram determinadas com base nas quantidades recuperadas. Assim, diferenças foram observadas entre as repetições. Incubou-se as três suspensões a 37°C por 3 h para estimular a liberação do conteúdo dos grânulos plaquetários..

## 2.6 Teste de *kill curve assay* (contagem de UFC/mL em placa de Petri)

O protocolo experimental de *kill curve assay* foi realizado de acordo com o proposto por Li [21]. Após crescimento overnight da bactéria *S. aureus* (MRSA e MSSA, em experimentos separados) em placa contendo meio de cultura ágar sangue ovino 5% (BPA-5%), foram adicionadas várias colônias bacterianas em 5 mL de caldo

Mueller-Hinton (MHB), a fim de se obter uma suspensão bacteriana, para cada bactéria, com densidade 0,5 na escala de McFarland (1-2 x 10<sup>8</sup> UFC/mL). O tubo foi homogeneizado vigorosamente em vortex e incubado a 37°C, 2h. A partir deste tubo, preparou-se uma diluição de 100x deste inóculo, utilizando-se solução salina tamponada de fosfatos (SST) para se obter a concentração de 1 x 10<sup>6</sup> UFC/mL. O inóculo foi mantido em gelo até o momento da inoculação nos tubos do experimento.

Microtubos 2,5 mL, esterilizados, com tampa, foram identificados de acordo com o quadro 1, e ensaiados como segue os mesmos quadros, respectivamente para cada bactéria, para um volume de 2 mL em cada tubo.

Primeiramente, foram adicionados PRP, WBC, P-PRP, ou SST aos respectivos microtubos, seguido da solução de ativação plaquetária (gluconato de cálcio, GC-10%) para ativação (formação de gel). Os microtubos foram incubados a 37°C com agitação orbital a 150 rpm em Agitador de Tubos de Ensaio 251 (Fanem, Brasil) para ativação plaquetária. Após, adicionou-se a MHB e, então, o inóculo bacteriano (1,0 x 10<sup>6</sup> UFC/mL) para obter uma concentração final de 1,0 x 10<sup>5</sup> UFC/mL. Após, os tubos foram incubados a 37°C com agitação orbital a 150 rpm em Agitador de Tubos de Ensaio 251 (Fanem, Brasil). Em momentos pré-determinados (*timepoints* 0 h, 1 h, 2 h), homogeneizou-se cada solução, em cada microtubo, por pipetagem repetitiva, pois a bactéria poderia estar presa no interior do gel. Pipetou-se 40 µL de amostra de cada tubo, diluindo-a seriadamente em solução salina estéril 0.9% (diluições 10-1, 10-2). Pipetou-se alíquota de 100 µL de cada diluição em ágar BPA-5%, em triplicata, para contagem de UFC/mL. As culturas foram incubadas a 37°C, overnight, e a contagem de UFC das placas realizada e registrada.

Quadro I - Experimento *kill curve assay* para avaliar a atividade antimicrobiana de PRP, WBC, e P-PRP junto as cepas de *Staphylococcus aureus* resistente à meticilina (MRSA - G1-G6) e *Staphylococcus aureus* sensível à meticilina (MRSA - G7-G12).

ID	Grupos	PRP / WBC / P-PRP	MHB	Bactéria	Suspensão bacteriana
G1A	Controle	-	1500 µL	MRSA	500 µL
G2A	Controle	-	2000 µL		-
G3A	Controle	SST + GC	q.s.p.		500 µL
G4A	PRP*	PRP + GC	q.s.p.		500 µL
G5A	WBC†	WBC + GC	q.s.p.		500 µL
G6A	P-PRP‡	P-PRP + GC	q.s.p.		500 µL
G7A	Controle	-	1500 µL	MSSA	500 µL
G8A	Controle	-	2000 µL		-

G9A	Controle	SST + GC	q.s.p.		500 µL
G10A	PRP*	PRP + GC	q.s.p.		500 µL
G11A	WBC†	WBC + GC	q.s.p.		500 µL
G12A	P-PRP‡	P-PRP + GC	q.s.p.		500 µL

\* q.s.p.: quantidade suficiente para completar 2000 µL

† PRP: concentração plaquetária de  $2,0 \times 10^6$  plaquetas/µL

‡ WBC: concentração leucocitária equivalente a contaminação leucocitária do PRP.

§ P-PRP: concentração plaquetária de  $2,0 \times 10^6$  plaquetas/µL

MHB: Mueller-Hinton

## 2.7 Análise estatística

Todos os dados referentes as contagens de unidades formadoras de colônia (UFC) foram tabulados em planilha Excel. Previamente as análises, foi verificada a distribuição dos dados pelo teste de normalidade de Kolmogorov-Smirnov (KS) para posterior seleção do melhor teste estatístico. Como os dados apresentaram distribuição normal, determinou-se a média e o erro-padrão da média (EPM) e aplicou-se o teste paramétrico de Análise de Variância para amostras dependentes (ANOVA), com o teste de Tukey para comparações múltiplas como pós-teste, para as análises das contagens de UFC dentro de um mesmo grupo, distribuídas nos três momentos analisados. Quando analisado o mesmo momento entre grupos diferentes, aplicou-se o teste ANOVA para amostras independentes. Para as comparações entre dois momentos pertencentes ao mesmo grupo, aplicou-se o teste t de Student (pareado), enquanto o teste t de Student (não pareado) foi aplicado para comparações a cada dois momentos pertencentes a grupos diferentes.

A redução ou aumento da multiplicação bacteriana nos *timepoints* 1 h e 2 h foi analisada com base nas médias dos valores obtidos nos *timepoints* prévios, obtidos respectivamente para cada bactéria, tomando-se como parâmetro basal os grupos controle basal para MRSA (G1) e MSSA (G7).

Para todas as análises considerou-se um nível de significância ( $\alpha$ ) de 5% [29]. Todas as análises foram realizadas nos programas GraphPad Prism v.5.01 e GraphPad Instat v.3.06 (GraphPad Software Inc. San Diego, CA, EUA).

## 3. RESULTADOS

Ao se avaliar o rendimento plaquetário pelas concentrações de plaquetas obtidas no PRP e P-PRP em relação àquela observada ao hemograma, verificou-se que o PRP rendeu uma média de 1,85x plaquetas a mais, enquanto 1,18x foi observada no P-PRP. Em termos absolutos, com base nos valores obtidos ao hemograma e mediante a quantidade de tubos coletados de cada animal para cada procedimento e volume de

sangue obtido, verificou-se que no PRP foi recuperada 15,42% das plaquetas que estavam nos tubos coletados, enquanto no P-PRP esse valor foi ligeiramente inferior (14,92%). A quantidade de plaquetas obtida no P-PRP foi 96,94% o valor obtido no PRP.

Ao se avaliar o grau de contaminação do PRP e P-PRP por leucócitos e eritrócitos, observou-se uma relação média de leucócitos:plaquetas de 1:12,59 e de eritrócitos:plaquetas de 21,25:1, enquanto no P-PRP não foi observada a presença de leucócitos, porém observou-se relação média de eritrócitos:plaquetas de 1:62,01.

Os resultados de média e erro-padrão da média para os grupos experimentais onde a bactéria MRSA foi desafiada aos diferentes protocolos de tratamento com plaquetas caninas em três diferentes momentos podem ser observados na Tabela 1 e na Figura 1. O único grupo referente a MRSA não plotado na figura, não presente na tabela e não analisado na estatística foi o G2 (controle negativo), pois não apresentou nenhum crescimento bacteriano.

Tabela I. Média e erro-padrão da média (EPM) para os grupos experimentais desafiados com a bactéria *Staphylococcus aureus* resistente a meticilina (MRSA) e submetidos ao tratamento com plaquetas caninas em três diferentes momentos. Presidente Prudente, 2021.

Momentos	Grupos experimentais												Valor de p	
	G1		G3		G4		G5		G6					
	Média	EPM	Média	EPM	Média	EPM	Média	EPM	Média	EPM				
0 hora	949,00 <sup>Aa</sup>	49,54	1103,17 <sup>Aab</sup>	59,58	966,00 <sup>Aab</sup>	98,54	1128,67 <sup>Aab</sup>	72,56	1245,50 <sup>Ab</sup>	63,59	0,0368			
1 hora	2713,00 <sup>Ba</sup>	275,54	4659,33 <sup>Bb</sup>	31,32	1704,50 <sup>ABa</sup>	479,95	1484,00 <sup>Aa</sup>	332,14	4785,33 <sup>Bb</sup>	232,15	<0.0001			
2 horas	6246,00 <sup>Cab</sup>	112,88	11372,00 <sup>Ca</sup>	78,08	4007,40 <sup>Bb</sup>	1364,17	3066,00 <sup>Ab</sup>	972,74	10166,00 <sup>Ca</sup>	630,00	0,0004			
<b>Valore de p</b>	< 0,0001		< 0,0001		0,0354		0,0799		< 0,0001					

Estatística: para um mesmo grupo experimental, médias de contagens de unidades formadoras de colônia (UFC) seguidas de letras maiúsculas diferentes indicam diferenças significativas entre os momentos analisados (*timepoints*), pela Análise de Variância para amostras dependentes, comparados pelo Teste de Tukey, enquanto que para um mesmo momento analisado, médias de contagens seguidas de letras minúsculas diferentes indicam diferenças significativas entre os grupos experimentais para aquele dado momento, pela Análise de Variância para amostras independentes, comparados pelo Teste de Tukey, considerando-se um nível de significância de 5%.

Legenda: EPM, erro-padrão da média; p, valor de p para  $\alpha = 0,05$ .

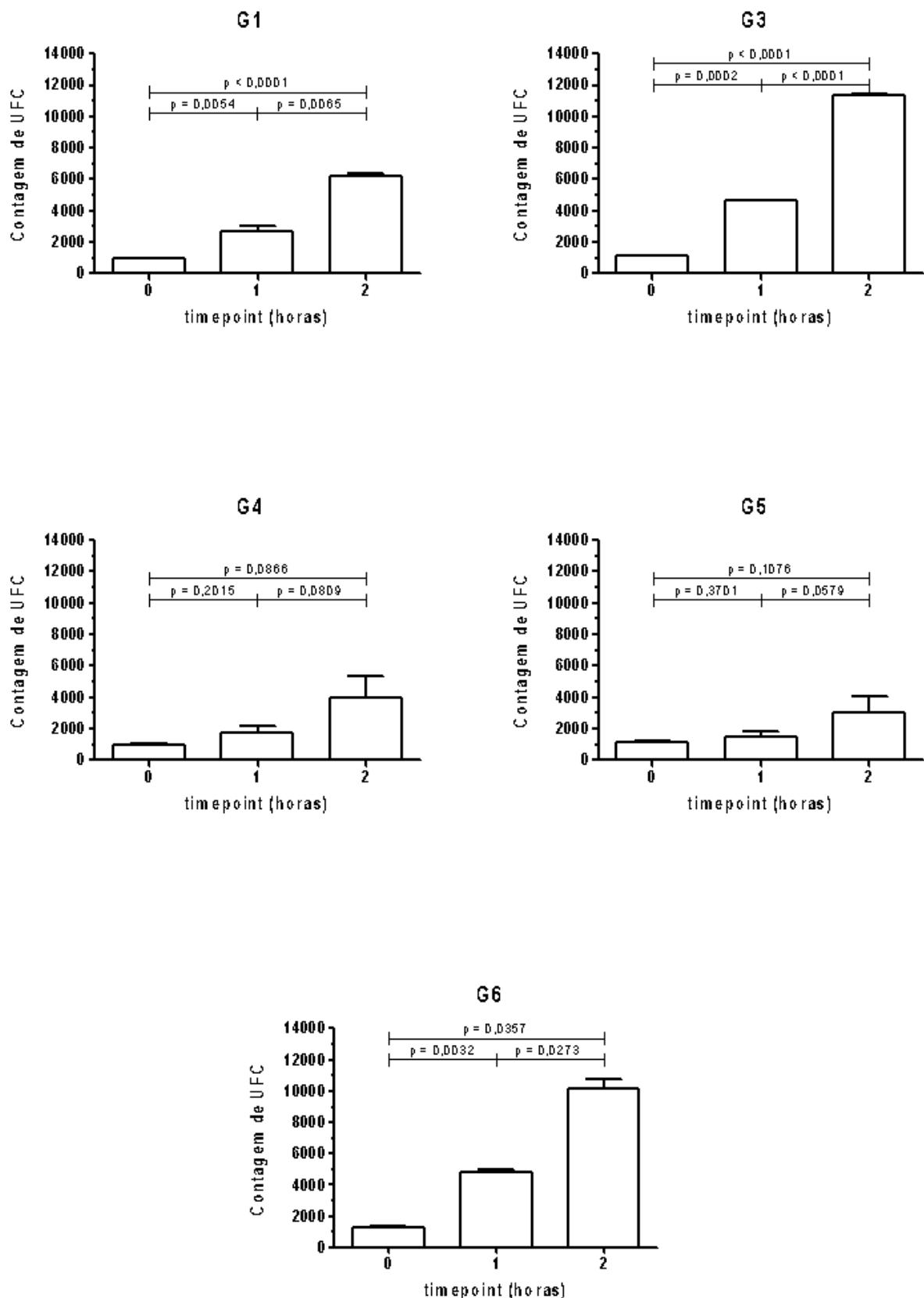


Figura I. Média e erro-padrão médio das concentrações de MRSA (UFC/mL) para os grupos G1-6 nos diferentes momentos de avaliação (*timepoints*).

Comparando-se os grupos experimentais, com ou sem tratamento com plaquetas, ao grupo controle basal (G1) do experimento com MRSA, a Figura 2 apresenta o efeito redutor ou amplificador das plaquetas e/ou leucócitos sobre a bactéria testada nas primeira e segunda horas testadas. Esta figura apresenta a porcentagem de redução ou amplificação da multiplicação bacteriana e a intensidade desta redução ou amplificação, sempre comparada ao grupo controle basal do experimento para MRSA.

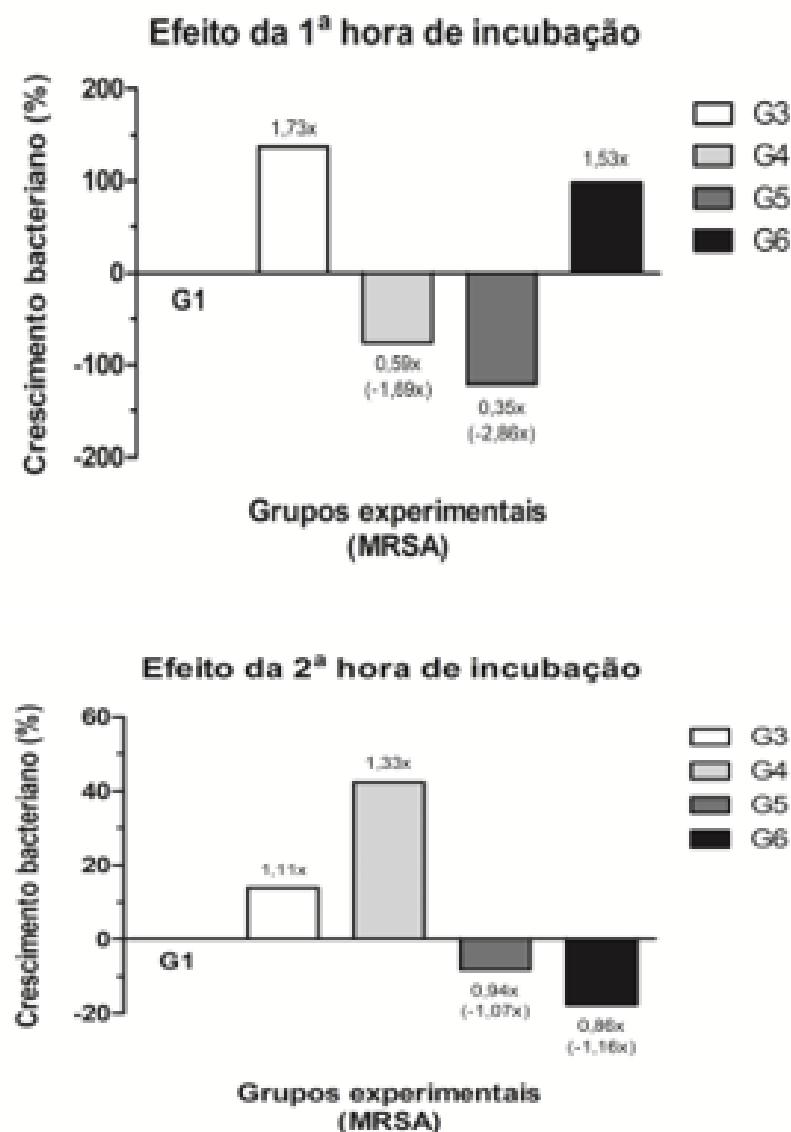


Figura II. Ação dos tratamentos sobre a multiplicação bacteriana nas primeira e segunda horas de incubação para *Staphylococcus aureus* resistente a meticilina (MRSA).

Legenda: sem parênteses estão presentes a relação de quantas vezes a porcentagem está em relação ao G1, isto é, em relação 0%. Valores acima de 1,0x indicam estímulo da multiplicação bacteriana, enquanto valores entre 0,0 e 1,0x indicam inibição ou redução da multiplicação bacteriana. Entre parênteses estão os valores inversamente proporcionais aos valores entre 0,0 e 1,0x, sendo os equivalentes de quantas vezes reduz a multiplicação bacteriana em relação ao G1.

Os resultados de média e erro-padrão da média para os grupos experimentais onde a bactéria MSSA foi desafiada aos diferentes protocolos de tratamento com plaquetas caninas em três diferentes momentos podem ser observados na Tabela 2 e na Figura 3. O único grupo referente a MSSA não plotado na figura, não presente na tabela e não analisado na estatística foi o G8 (controle negativo), pois não apresentou nenhum crescimento bacteriano.

Tabela II. Média e erro-padrão da média (EPM) para os grupos experimentais desafiados com a bactéria *Staphylococcus aureus* sensível a meticilina (MSSA) e submetidos ao tratamento com plaquetas caninas em três diferentes momentos. Presidente Prudente, 2021.

Momentos	Grupos experimentais										Valor de p	
	G7		G9		G10		G11		G12			
	Média	EPM	Média	EPM	Média	EPM	Média	EPM	Média	EPM		
0 hora	882,33 <sup>Aa</sup>	5,36	906,33 <sup>Aa</sup>	14,50	849,67 <sup>Aa</sup>	11,41	661,00 <sup>Ab</sup>	30,62	782,00 <sup>Aa</sup>	43,00	0,0003	
1 hora	1777,67 <sup>Ba</sup>	18,78	2095,00 <sup>Bb</sup>	293,86	632,00 <sup>Ac</sup>	85,54	636,00 <sup>Ac</sup>	87,27	1767,00 <sup>Ba</sup>	35,55	<0.0001	
2 horas	4074,67 <sup>Ca</sup>	247,60	6289,67 <sup>Cb</sup>	432,15	719,00 <sup>Ac</sup>	84,79	983,67 <sup>Ac</sup>	107,42	5189,67 <sup>Cab</sup>	367,16	<0.0001	
<b>Valore de p</b>	0,0002		< 0,0001		0,2646		0,1042		<0,0001			

Estatística: para um mesmo grupo experimental, médias de contagens de unidades formadoras de colônia (UFC) seguidas de letras maiúsculas diferentes indicam diferenças significativas entre os momentos analisados (*timepoints*), pela Análise de Variância para amostras dependentes, utilizando o Teste de Tukey como pós-teste, enquanto que para um mesmo momento analisado, médias de contagens seguidas de letras minúsculas diferentes indicam diferenças significativas entre os grupos experimentais para aquele dado momento, pela Análise de Variância para amostras independentes, também utilizando o Teste de Tukey como pós-teste, considerando-se um nível de significância de 5%.

Legenda: EPM, erro-padrão da média; p, valor de p para  $\alpha = 0,05$ .

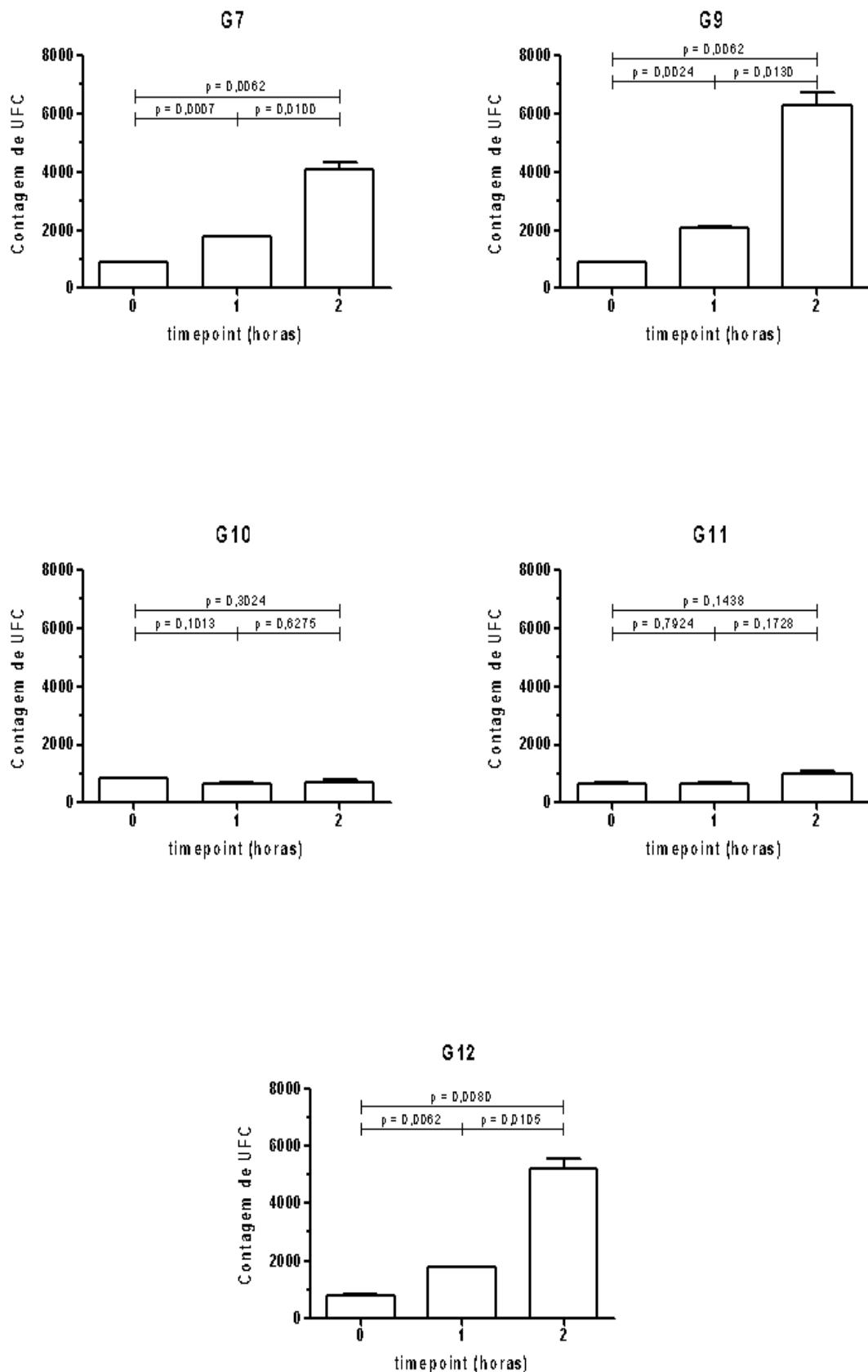


Figura III. Média e erro-padrão médio das concentrações de MSSA (UFC/mL) para os grupos G7-12 nos diferentes momentos de avaliação (*timepoints*).

Comparando-se os grupos experimentais, com ou sem tratamento com plaquetas, ao grupo controle basal (G7) do experimento com MSSA, a Figura 4 apresenta o efeito redutor ou amplificador das plaquetas e/ou leucócitos sobre a bactéria testada nas primeira e segunda horas testadas. Esta figura apresenta a porcentagem de redução ou amplificação da multiplicação bacteriana e a intensidade desta redução ou amplificação, sempre comparada ao grupo controle basal do experimento para MSSA.

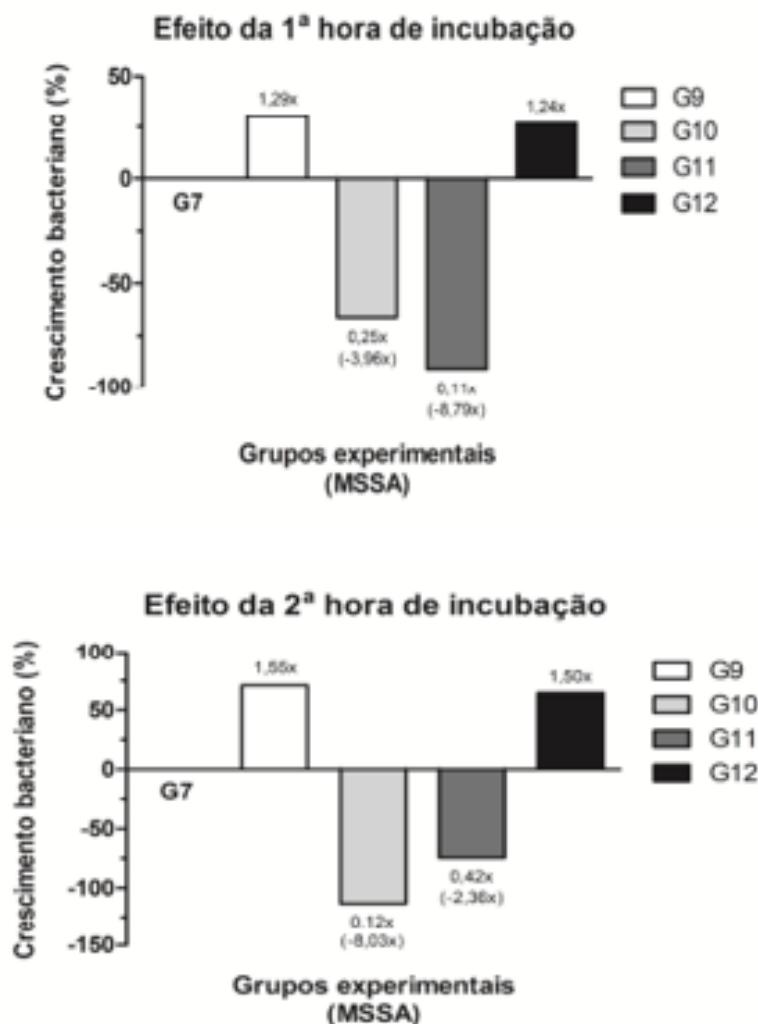


Figura IV. Ação dos tratamentos sobre a multiplicação bacteriana nas primeira e segunda horas de incubação para *Staphylococcus aureus* sensível a meticilina (MSSA). Legenda: sem parênteses estão presentes a relação de quantas vezes a porcentagem está em relação ao G7, isto é, em relação 0%. Valores acima de 1,0x indicam estímulo da multiplicação bacteriana, enquanto valores entre 0,0 e 1,0x indicam inibição ou redução da multiplicação bacteriana. Entre parênteses estão os valores inversamente proporcionais aos valores entre 0,0 e 1,0x, sendo os equivalentes de quantas vezes reduz a multiplicação bacteriana em relação ao G7.

#### 4. DISCUSSÃO

Na comparação entre momentos no mesmo grupo houve aumento ( $P<0,05$ ) no número de UFCs ao longo do tempo de avaliação em todos os grupos, porém sem significância estatística para os grupos G4( $P=0,0354$ ) e G5 ( $P=0,0799$ ) onde os valores permaneceram semelhantes ao momento 0h até a primeira hora para ambos os grupos e até duas horas para o G5, mostrando neste grupo o efeito máximo na inibição do crescimento de MRSA na primeira hora. Nos diferentes grupos, no *timepoint* 0h houve aumento das UFCs no G6 diferindo significativamente do G1 ( $P=0,0368$ ); com 1h ( $P<0,0001$ ) e 2h ( $P=0,0004$ ) já os grupos G4 (PRP) e G5 (WBC) se mantiveram com valores semelhantes ao controle e significativamente menores em relação aos grupos G3 e G6. Os resultados para os grupos experimentais desafiados com a bactéria *Staphylococcus aureus* sensível a meticilina (MSSA) e submetidos ao tratamento com plaquetas caninas em três diferentes momentos, mostram o aumento significativo nas UFCs ao longo do tempo de avaliação em todos os grupos, no entanto, para os grupos G10( $P=0,2646$ ) e G11( $P=0,1042$ ) não houve diferença ao longo do tempo e os menores valores para as UFCs ocorreram na primeira hora para estes dois grupos, sendo valores inferiores aos do momento 0h. Na comparação entre grupos nos diferentes momentos no *timepoint* 0h houve redução das UFCs no grupo G11 (WBC) diferindo significativamente ( $P=0,0003$ ) dos demais grupos. Com 1h e 2h a redução ocorreu não só no grupo G11 mas também no grupo G10 diferindo significativamente ( $P<0,0001$ ) dos demais grupos, desta forma os achados do presente estudo mostram a ação positiva do PRP e WBC como já citado por outros autores [30-32].

Em situações em que existe uma solução de continuidade, que pode ocorrer por diferentes motivos, pode existir comprometimento além da pele, do tecido celular subcutâneo e muscular e em casos de colonização bacteriana destas lesões, o processo de reparação e cicatrização pode ficar prejudicado e retardado. Na literatura existem evidências de que o concentrado de plaquetas, além de efeito regenerativo, tem efeito antimicrobiano por ação direta no reconhecimento, sequestro e neutralização de patógenos invasores e indireta no recrutamento e modulação de leucócitos para locais de infecção proporcionando maior capacidade fagocitária devido as diferentes vias de sinalização, o que pode ajudar o processo de cicatrização [11-20-32-33].

No presente estudo, o efeito bacteriostático sobre a atividade de MRSA e MSSA foi confirmado nos grupos G4, G5 e G10, G11 respectivamente, corroborando outros autores [31-32]. Além disso, foi observado que a concentração leucocitária do PRP foi o que mais interferiu para redução das UFCs, sendo que para MSSA, a inibição foi ainda mais acentuada quando comparado a MRSA, achado relatado por outros autores em situações clínicas de redução de infecção em casos ortopédicos [34], trauma [35], cirurgia maxilo-facial [32] e cardíaca [36].

A redução das UFCs nos grupos G4 e G10 se relacionam com as atividades antimicrobianas das plaquetas, que promovem um papel defensivo natural podendo estar diretamente ligados à presença de leucócitos contidos no biomaterial que pode levar a degranulação das memas. Os grânulos alfa ricos e fatores de crescimento, citocinas, proteínas/peptídeos microbicidas plaquetários (PMPs) contidos no interior das plaquetas funcionam como quimiocinas, podendo ser usada como terapia complementar na infecção de feridas, uma vez que possui um menor risco de resistência quando comparada aos fármacos ou ajudando para feridas crônicas não infectadas sejam colonizadas [19-37-38].

Apesar de ser observado o efeito bacteriostático do PRP neste estudo, também relatado por Smith et al. [39] frente a *S. aureus* independentemente do PRP estar ativado ou inativado, o maior efeito inibitório sobre o crescimento bacteriano se deu nos grupos G5 e G11 e estão relacionados ao importante papel dos leucócitos, porém nos grupos G4 e G10 observamos o importante sinergismo das plaquetas com os leucócitos, estes podem ter seu número reduzido ou aumentado durante o processamento do PRP [40], portanto devemos levar em consideração que fatores como diferentes técnicas [41], método de ativação exógena [42] e variabilidade biológica da concentração do fator de crescimento entre os indivíduos [43] podem modificar a composição do PRP e a quantidade de fatores de crescimento disponíveis. Neste estudo, produzimos e utilizamos uma contagem de plaquetas seis vezes maior que a contagem inicial e um aumento de três a cinco vezes nas plaquetas, o que tem sido considerado uma concentração apropriada para aplicações médicas [44].

A agregação leucócito-plaquetas causa uma resposta inflamatória aumentada, disponibilizando um maior número de fator de crescimento para tecidos lesionados comparado com métodos pobres em leucócitos [45], além de promover a liberação de serino-proteases e metaloproteinases (MMP) que controlam a resposta inflamatória, no entanto, também liberam citocinas inflamatórias e espécies reativas de

oxigênio que são prejudiciais durante o tratamento [46-47]. Desta forma, existe uma controvérsia com relação a concentração de leucócitos que seria ideal e em que situação clínica seu uso seria indicado, apesar da confirmação em estudo de revisão sistemática, de que o concentrado de plaquetas possui efeito antimicrobiano [48]. De qualquer forma, sugerimos que outros estudos com diferentes concentrações leucocitárias no PRP sejam realizados para determinar seu envolvimento sobre o efeito bacteriostático e as diferentes formulações sejam testadas *in vivo*.

## 5. CONCLUSÃO

Conclui-se que as plaquetas desempenham papel importante na inibição/redução sendo potentes moduladoras e efetoras imunológicas na multiplicação bacteriana *in vitro* das bactérias MRSA e MSSA, efeito semelhante também ocorre quando na presença isolada de leucócitos (WBC), porém observamos que ocorre uma interação sinérgica entre plaqueta-leucócitos-patógeno no PRP principalmente nos primeiros momentos de exposição ao patógeno, no entanto o P-PRP estimula o crescimento bacteriano, assim entender os mecanismos envolvidos é fundamental para elaboração de abordagens e protocolos terapêuticos eficazes.

## 6. AGRADECIMENTOS

À CAPES pelo incentivo financeiro concedido: “O presente trabalho foi realizado com apoio da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – (Brasil) CAPES

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## ANEXO A- APROVAÇÃO ÉTICA

### UNOESTE - Universidade do Oeste Paulista

PRO-REITORIA DE PESQUISA E POS-GRADUAÇÃO

PPG - Programa de Pesquisa de Pós-Graduação  
PEIC - Programa Especial de Iniciação Científica

### Parecer Final

Declaro para os devidos fins que o Projeto de Pesquisa intitulado "EFEITO BACTERICIDA DO PLASMA RICO EM PLAQUETAS (PRP) EM BACTÉRIAS STAPHYLOCOCCUS AUREUS SENSÍVEL A METICILINA (MSSA) E STAPHYLOCOCCUS AUREUS RESISTENTES A METICILINA (MRSA): ESTUDO IN VITRO UTILIZANDO TESTE KILL CURVE ASSAY", cadastrado na Coordenadoria de Pesquisa, Desenvolvimento e Inovação (CEDI) sob o número nº 5969 e tendo como participante(s) ANA FLÁVIA SORIANO PEREIRA (discente), ANDERSON MAGALHÃES (discente), CECILIA LAPOSY SANTAREM (docente), RODRIGO COSTA DA SILVA (docente), ROSA MARIA BARILLI NOGUEIRA (orientador responsável), foi avaliado e APROVADO pelo COMITÊ ASSESSOR DE PESQUISA INSTITUCIONAL (CAPI) e COMISSÃO DE ÉTICA USO DE ANIMAIS (CEUA) da Universidade do Oeste Paulista - UNOESTE de Presidente Prudente/SP.

Este Projeto de Pesquisa, que envolve a produção, manutenção e/ou utilização de animais pertencentes ao filo Chordata, subfilo Vertebrata (exceto o homem), para fins de pesquisa científica, encontra-se de acordo com os preceitos da Lei nº 11.794, de 8 de Outubro de 2008, do Decreto nº 6.899, de 15 de Julho de 2009, e com as normas editadas pelo Conselho Nacional de Controle da Experimentação Animal (CONCEA), tendo sido APROVADO em reunião realizada em 12/02/2020.

Vigência do projeto: 12/2019 a 04/2022.

#### ANIMAL VIVO

Espécie/Linagem	Nº de Animais	Peso	Idade	Sexo	Origem
c	4	25 quilos	5 anos	F	Canil Unoeste

Presidente Prudente, 24 de Fevereiro de 2020.

Prof. Dr. Adilson Rodrigues Garcia Jr.  
Coordenador Científico da CEDI

Prof. Ms. Adriana Fábio de Brito  
Coordenadora da CELIA - UNOESTE

Coordenadoria de Pesquisa, Desenvolvimento e Inovação - CEDI - 18.2020-2079 - [capi@unesp.br](mailto:capi@unesp.br)  
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## **ANEXO B– NORMAS PARA AUTORES DA REVISTA: BRAZILIAN JOURNAL OF MICROBIOLOGY**

### **Type of Articles**

The Brazilian Journal of Microbiology accepts submissions of the following article types:

Research Papers: report results of original research, which has not been published elsewhere. Short communications: a short communication should report new and significant findings. Submit form is the same way as research paper. They receive the same review, they are not published more rapidly than research paper.

Reviews: Review articles should deal with microbiological subjects of broad interest.

Letters to the editor: letters to the editor are intended only for comments on final, typeset articles published in the journal (manuscripts posted online are not accepted) and must cite published references to support the writer's argument.

Your manuscript must be written clearly, in comprehensible and linguistically correct English. Manuscripts written in poor English will not be accepted. Please check the section "English Language Support" how to get assistance.

**Sections** The Brazilian Journal of Microbiology has the following sections (one of them should be selected during the electronic submission process):

Biotechnology and Industrial Microbiology: Biosynthesis and bioconversion of natural products, including antibiotics, xenobiotics, and macromolecules produced by bacteria.

Biosynthesis and bioconversion of natural products, including antibiotics, xenobiotics, and macromolecules produced by fungi. Molecular aspects of fungal biotechnology.

Molecular aspects of bacterial biotechnology.

Food Microbiology: Applications of microorganisms (bacteria and fungi) for food production. Food borne diseases, food spoilage, and microbial ecology in foods.

Bacterial and Fungal Pathogenesis: The genetic, biochemical, and structural basis of bacterial pathogenesis.

Clinical Microbiology: Studies of medically-important bacteria, fungi and virus.

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The trial registration number (TRN) and date of registration should be included as the last line of the manuscript abstract.

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Purely observational trials will not require registration.

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Springer Nature advocates complete and transparent reporting of biomedical and biological research and research with biological applications. Authors are recommended to adhere to the minimum reporting guidelines hosted by the EQUATOR Network when preparing their manuscript.

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#### Summary of requirements

The above should be summarized in a statement and included on a title page that is separate from the manuscript with a section entitled "Declarations" when submitting a paper. Having all statements in one place allows for a consistent and unified review of the information by the Editor-in-Chief and/or peer reviewers and may speed up the handling of the paper. Declarations include Funding, Conflicts of interest/competing interests, Ethics approval, Consent, Data and/or Code availability and Authors' contribution statements. Please use the following template title page for providing the statements.

Once and if the paper is accepted for publication, the production department will put the respective statements in a distinctly identified section clearly visible for readers.

Please see the various examples of wording below and revise/customize the sample statements according to your own needs.

- Provide "Ethics approval" as a heading (see template)

Examples of ethics approval obtained:

- All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. The study was approved by the Bioethics Committee of the Medical University of A (No. ...).

- This study was performed in line with the principles of the Declaration of Helsinki. Approval was granted by the Ethics Committee of University B (Date.../No. ...).

- Approval was obtained from the ethics committee of University C. The procedures used in this study adhere to the tenets of the Declaration of Helsinki.

- The questionnaire and methodology for this study was approved by the Human Research Ethics committee of the University of C (Ethics approval number: ...).

Examples of a retrospective study:

- Ethical approval was waived by the local Ethics Committee of University A in view of the retrospective nature of the study and all the procedures being performed were part of the routine care.
- This research study was conducted retrospectively from data obtained for clinical purposes. We consulted extensively with the IRB of XYZ who determined that our study did not need ethical approval. An IRB official waiver of ethical approval was granted from the IRB of XYZ.
- This retrospective chart review study involving human participants was in accordance with the ethical standards of the institutional and national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. The Human Investigation Committee (IRB) of University B approved this study.

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- This is an observational study. The XYZ Research Ethics Committee has confirmed that no ethical approval is required.
- The data reproduced from Article X utilized human tissue that was procured via our Biobank AB, which provides de-identified samples. This study was reviewed and deemed exempt by our XYZ Institutional

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Identifying details (names, dates of birth, identity numbers, biometrical characteristics (such as facial features, fingerprint, writing style, voice pattern, DNA or other distinguishing characteristic) and other information) of the participants that were studied should not be published in written descriptions, photographs, and genetic profiles unless the information is essential for scholarly purposes and the participant (or parent or guardian if the participant is incapable) gave written informed consent for publication. Complete anonymity is difficult to achieve in some cases. Detailed descriptions of individual participants, whether of their whole bodies or of body sections, may lead to disclosure of their identity. Under certain circumstances consent is not required as long as information is anonymized and the submission does not include images that may identify the person.

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When biological material is donated for or data is generated as part of a research project authors should ensure, as part of the informed consent procedure, that the participants are made what kind of (personal) data will be processed, how it will be used and for what purpose. In case of data acquired via a biobank/biorepository, it is possible they apply a broad consent which allows research participants to consent to a broad range of uses of their data and samples which is regarded by research ethics committees as specific enough to be considered “informed”. However, authors should always check the specific biobank/biorepository policies or any other type of data provider policies (in case of non-bio research) to be sure that this is the case.

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For all research involving human subjects, freely-given, informed consent to participate in the study must be obtained from participants (or their parent or legal guardian in the case of children under 16) and a statement to this effect should appear in the manuscript. In the case of articles describing human transplantation studies, authors must include a statement declaring that no organs/tissues were obtained from prisoners and must also name the institution(s)/clinic(s)/department(s) via which organs/tissues were obtained. For manuscripts reporting studies involving vulnerable groups where there is the potential for coercion or where consent may not have been fully informed, extra care will be taken by the editor and may be referred to the Springer Nature Research Integrity Group.

Consent to Publish

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Summary of requirements

The above should be summarized in a statement and included on a title page that is separate from the manuscript with a section entitled “Declarations” when submitting a paper. Having all statements in one place allows for a consistent and unified review of the information by the Editor-in-Chief and/or peer reviewers and may speed up the handling of the paper. Declarations include Funding, Conflicts of interest/competing interests, Ethics approval, Consent, Data and/or Code availability and Authors’ contribution statements. Please use the template Title Page for providing the statements. Once and if the paper is accepted for publication, the production department will put the respective statements in a distinctly identified section clearly visible for readers.

Please see the various examples of wording below and revise/customize the sample statements according to your own needs.

Provide “Consent to participate” as a heading

Sample statements consent to participate:

- Informed consent was obtained from all individual participants included in the study.
- Informed consent was obtained from legal guardians.
- Written informed consent was obtained from the parents.
- Verbal informed consent was obtained prior to the interview.
- The patient has consented to the submission of the case report for submission to the journal.

Provide “Consent to publish” as a heading

- The authors affirm that human research participants provided informed consent for publication of the images in Figure(s) 1a, 1b and 1c.
- The participant has consented to the submission of the case report to the journal.
- Patients signed informed consent regarding publishing their data and photographs.
- Sample statements if identifying information about participants is available in the article:
- Additional informed consent was obtained from all individual participants for whom identifying information is included in this article.
- Additional informed consent was obtained from all individual participants for whom identifying information is included in this article.
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### **Research involving animals**

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ICMJE, Defining the Role of Authors and Contributors,  
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scientific publication, McNutt et al, PNAS February 27, 2018

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All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by [full name], [full name] and [full name]. The first draft of the manuscript was written by [full name] and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

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A Graduate Student's Guide to Determining Authorship Credit and Authorship Order, APA Science Student Council 2006

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